

RESEARCH ARTICLE

Subgingival areas as potential reservoirs of different *Candida* spp in type 2 diabetes patients and healthy subjects

Sanja Matic Petrovic¹✉, Milena Radunovic²✉*, Milena Barac¹, Jovana Kuzmanovic Pfcic³, Dusan Pavlica², Valentina Arsic Arsenijevic⁴, Ana Pucar¹*

1 Department of Oral Medicine and Periodontology, School of Dental Medicine, University of Belgrade, Belgrade, Serbia, **2** Department of Microbiology, School of Dental Medicine, University of Belgrade, Belgrade, Serbia, **3** Department for Medical Statistics and Informatics, School of Dental Medicine, University of Belgrade, Belgrade, Serbia, **4** Department of Microbiology and Immunology, School of Medicine, University of Belgrade, Belgrade, Serbia

✉ These authors contributed equally to this work.
* anapucar@gmail.com (AP); milena.radunovic@stomf.bg.ac.rs (MR)



OPEN ACCESS

Citation: Matic Petrovic S, Radunovic M, Barac M, Kuzmanovic Pfcic J, Pavlica D, Arsic Arsenijevic V, et al. (2019) Subgingival areas as potential reservoirs of different *Candida* spp in type 2 diabetes patients and healthy subjects. PLoS ONE 14(1): e0210527. <https://doi.org/10.1371/journal.pone.0210527>

Editor: David M. Ojcius, University of the Pacific, UNITED STATES

Received: October 9, 2018

Accepted: December 26, 2018

Published: January 10, 2019

Copyright: © 2019 Matic Petrovic et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its Supporting Information files.

Funding: This work has been financed by the Grants No 41008 and 41027 of the Ministry of Education, Science and Technological Development of Serbia.

Competing interests: The authors have declared that no competing interests exist.

Abstract

Objectives

The aim of this cross-sectional observational study was to compare the prevalence of different oral *Candida* spp. in patients with Type 2 Diabetes and chronic periodontitis in two oral sites: dorsal surface of the tongue and subgingival area. In order to determine subgingival areas as potential reservoirs of yeasts, this study aimed to find differences in the yeasts' detection between the dorsum of the tongue, as the oral site most commonly inhabited with microorganisms, and subgingival samples. Additionally, potential predictors for the yeasts prevalence were determined.

Material and methods

Subjects (N = 146) were divided into four groups: group A- healthy individuals without periodontitis, group B- healthy individuals with chronic periodontitis, group C- Type 2 Diabetes patients with good glycoregulation and Chronic periodontitis and group D- Type 2 Diabetes patients with poor glycoregulation and Chronic periodontitis. Samples were obtained from the tongue by swabbing. Subgingival plaque samples were taken by paper points and periodontal curette. Isolation and identification of different *Candida* spp. was done using ChromAgar medium. In addition, germ-tube production and carbohydrate assimilation tests were performed.

Results

The prevalence of *Candida* spp. was higher in diabetics with poor glycoregulation. The most frequently isolated species was *Candida albicans* followed by *Candida glabrata* and *Candida tropicalis*. In 15.6% of cases, *Candida* spp. was present in the subgingival area while absent on the tongue. Multivariate regression model showed that HbA1c was *Candida* spp. predictor for both locations.

Conclusions

Our results confirmed that there are *Candida* spp. carriers among subjects with clinically healthy oral mucosa. Also, this study identified subgingival areas as potential reservoirs of these pathogenic species. Glycoregulation has been recognized as a positive predictor factor of *Candida* spp.

Introduction

Chronic periodontitis (CP) and oral mucosal candidiasis have been considered as chronic complications of Type 2 Diabetes (T2D) [1, 2]. Immunocompromised patients (e.g. Diabetics, AIDS patients) are more prone to *Candida* colonization and infection [1, 3–5]. In the oral cavity, *Candida* spp. is most commonly found on the dorsal surface of the tongue, followed by the palatal and buccal mucosa [6]. However, yeasts have also been found in subgingival areas [7]. Bacteria in the dental biofilm are thought to be the main etiological factor of periodontitis, but there is an growing evidence about the importance of viruses [8] and yeasts [9] in pathogenesis of chronic periodontitis. There is still a disagreement if the yeasts are transient members of oral biofilms [10], or if they are definite members of the oral microbiome [11]. Subgingival yeasts are present in 10–30% of healthy subjects [11, 12] and in up to 52% of diabetics [11, 13, 14].

Although *Candida albicans* is the most frequent species of *Candida* genus, the prevalence and importance of other species, such as *Candida glabrata*, *Candida tropicalis*, *Candida krusei* and *Candida dubliniensis* are increasing [4]. These, non-*albicans* species were first considered as markers of immunocompromised subjects, but have recently also been isolated from healthy subjects [3, 11].

Since antibiotics are commonly used in the treatment of periodontitis, allowing yeasts to proliferate freely, the possible role of yeasts in the pathogenesis of periodontal diseases is important, especially for immunocompromised subjects.

The purpose of this study was to compare the frequency of different *Candida* spp. on the tongue and in the subgingival sites between healthy subjects and T2D patients with chronic periodontitis. In order to determine subgingival areas as potential reservoirs of yeasts, this study aimed to find differences in the yeasts' detection between the dorsum of the tongue, as the oral site most commonly inhabited with microorganisms, and subgingival samples. Additionally, potential predictors for the yeasts prevalence were determined.

Material and methods

Study population and study design

This cross-sectional observational study, conducted from February 2015 to April 2017, has the approval of Ethical Committee of the School of Dental Medicine, University of Belgrade in accordance with the Declaration of Helsinki (Ethics Approval no. 36/8, 20 Feb 2015). A total of 146 subjects, who signed an informed consent, were selected and classified in four -experimental groups (Fig 1):

- Group A: 36 healthy volunteers without clinical signs of periodontitis (Probing pocket depth, PPD < 3 mm, Clinical Attachment Loss, CAL = 0 mm, Bleeding on Probing, BOP < 25%)

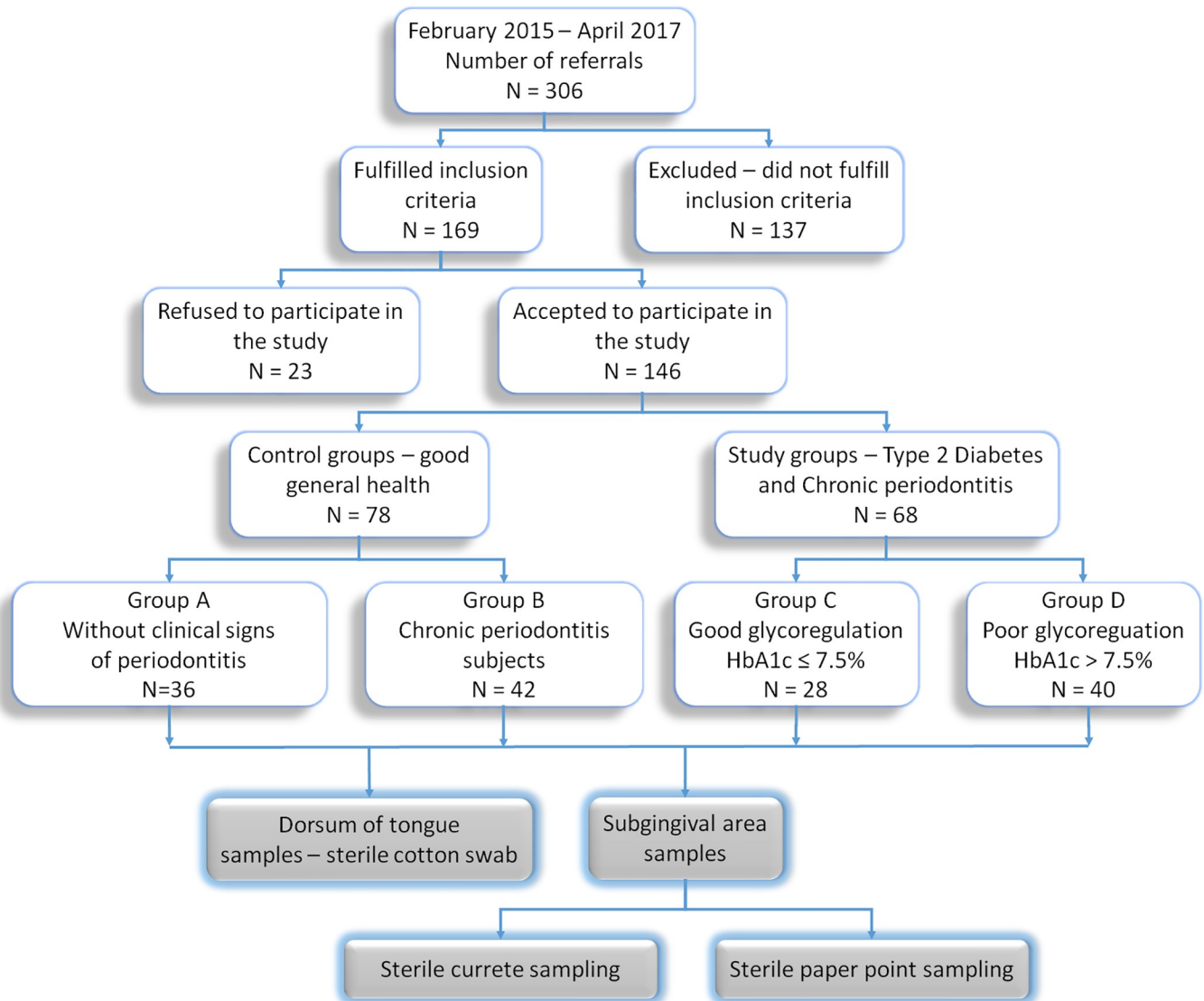


Fig 1. Study and subject design.

<https://doi.org/10.1371/journal.pone.0210527.g001>

- Group B: 42 patients with diagnosed CP and free of systemic diseases. Generalized CP was diagnosed if the subject exhibited $CAL \geq 1$ mm and $PPD > 3$ mm at $>30\%$ of sites. According to CAL, periodontitis was defined as slight ($CAL = 1-2$ mm), moderate ($CAL = 3-4$ mm) or severe ($CAL > 5$ mm) [15]. Patients with periodontitis were referred to the Department of Periodontology and Oral Medicine, School of Dental Medicine, University of Belgrade.
- Group C: 28 patients with diagnosed CP and T2D with satisfactory metabolic control ($HbA_{1c} \leq 7.5\%$). T2D was diagnosed by measuring glycated hemoglobin (HbA_{1c}) and blood glucose levels (using oral glucose tolerance test) [16].
- Group D: 40 patients with diagnosed CP, T2D and with poor metabolic control ($HbA_{1c} > 7.5\%$). Diabetic patients (C and D groups) were referred to the Clinic for Endocrinology, Diabetes and Metabolic Diseases, Clinical Center of Serbia.

Inclusion and exclusion criteria

In order to take part in the study, the subjects were expected to have more than 14 teeth, and not show any of the following traits: presence of any disease except T2D and its complications, aggressive periodontitis, therapy that could affect the periodontium (antibiotics, topical or systematic corticosteroids or antiseptics), previous oral candidiasis treatment, pregnancy/lactation and periodontal treatment in the last 1.5 year.

Socio-demographic and lifestyle data, clinical and microbiological outcomes and additional measures

Each patient had undergone 1) socio-demographic and lifestyle data using questionnaire interview; 2) clinical evaluation and measurements (secondary outcomes); 3) microbiological sampling procedures (primary outcomes) and 4) additional measurements.

1) Socio-demographic and lifestyle data using questionnaire interview included age, gender and education data. Also, parameters that could potentially influence clinical parameters of periodontitis and/or T2D or *Candida* carriage, but which could not be regarded as exclusion criteria were recorded: self-reported information about xerostomia, blood type, everyday intake of sweets and smoking habits (“non-smokers” and “smokers”). Patients who ceased smoking less than 6 months prior to the study were not included

2) Clinical evaluation and measurement (a full mouth clinical examination) was performed by two experienced and calibrated doctors, using Williams’ probe calibrated in millimeters (Hu-Friedy Chicago, IL). The following periodontal parameters at six sites per tooth were assessed: 1) plaque index-Silness Loe (PI), index of oral hygiene based on recording both soft debris and mineralized deposits on tooth, 2) bleeding on probing (BOP), measured 15 seconds after probing, defined as presence or absence, 3) probing pocket depth (PPD) as a distance from a margin of gingiva to the bottom of the sulcus/pocket (expressed in mm), 4) clinical attachment level (CAL) as distance from cemento-enamel junction to the pocket bottom (expressed in mm). The PPD and CAL were rounded to the higher millimeter and done at all teeth except third molars. For the evaluation of intra-examiner agreement, 20% of samples were reexamined 2 days after the first examinations.

3) Microbiological sampling procedures started a day after periodontal examination. Oral swabs were collected using a sterile cotton stick by swabbing ten times from the dorsum of the tongue and immediately inoculated on Sabouraud dextrose agar (Oxoid, Basingstoke, UK). The subgingival samples were obtained from the tooth with deepest PPD. First, cotton rolls were used to isolate the tooth and the supragingival plaque was removed by a curette and sterile gauze. Two sterile paper points (#30) were placed into the pocket/sulcus until a mild resistance appeared and kept for 30 seconds. Paper points contaminated by blood were excluded. Immediately after, sterile curette (S4L/4R SS G.Hartzell&Son, Concord, California) was used to obtain samples of the complete subgingival biofilm. Both subgingival samples were inoculated in separate sterile plastic tubes containing 1 mL of Sabouraud dextrose broth, vortexed for 60 seconds. From each sample, 20µl of suspended broth was streaked on SDA using sterile plastic micro pipette and incubated at 37°C for 48 hours. The number of Colony Forming Units per milliliter (CFU/ml) for samples taken by paper points was counted by one calibrated microbiologist. Detailed protocol can be found at <http://dx.doi.org/10.17504/protocols.io.wdefa3e>. The same microbiologist defined growth for samples taken from tongue as rare (number of CFU per sample ≤ 5), medium (6–50 CFU per sample) and dense (number of CFU per sample > 50).

Distinction of different species of *Candida* genus was done using CHROMagar *Candida* Medium (Becton Dickinson) according to producers instructions, germ-tube production test

and carbohydrate assimilation test [17]. After identification of more than one *Candida* species per sample, the levels of each species were defined as percentage of overall culture growth. The microbiologist was blinded for any clinical and anamnestic information.

After cultivation or second clinical examination (for examination of intra rater reliability), subjects received necessary dental treatment.

4) Additional measurements include biochemical and hematological analysis: fasting plasma glucose levels (FPG), HbA1c, hematological parameters: red blood count (RBC), hemoglobin (Hgb), hematocrit (HCT), mean corpuscular volume (MCV), mean cell hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC) and sedimentation rate.

Statistical analysis

SPSS 22.0 software package for Windows (SPSS inc. Chicago, USA) was used for statistical analysis. Descriptive data were presented as Mean (SD) for numerical or the percentage for discrete measures. ANOVA with Bonferroni correction was used for normally distributed data. Non-parametric data was analyzed using the Kruskal-Wallis and Mann-Whitney test. Chi Square Test (χ^2) was used for comparison of categorical variables. The logistic regression model was used to determine predictors of the presence of *Candida* spp. Subjects with missing data were not included in the study. Inter-rater reliability was appraised for each periodontal clinical parameter using the Cohen's kappa statistics. Differences were considered significant when p-value was < 0.05 .

Results

Socio-demographic, clinical outcome variables and additional measurements

According to results from questionnaires, we found that groups were similar by age, gender and smoking habits (Table 1). Also, subjects with O vs. A/AB/B blood type system were equally distributed between groups (Table 1).

Clinical examination showed that groups differed according to the number of present teeth (Table 1). Intergroup analysis showed that similar number of teeth was found only between groups C and D (Mann-Whitney U Test, $p = 0.073$), while other groups exhibited different number of teeth (Mann-Whitney U Test, A vs. B: $p < 0.0001$, A vs. C: $p < 0.0001$, A vs. D: $p < 0.0001$, B vs. C: $p = 0.009$, B vs. D: $p < 0.0001$). PI was the highest in diabetics with good glycorregulation, and differed between all groups (Mann-Whitney U test, $p < 0.05$ for all intergroup comparisons). BOP was significantly higher in groups B, C and D than in group A (ANOVA with Bonferroni correction, $p = 0.001$, $p = 0.002$ and $p < 0.0001$, respectively). PPD differed between A vs. B, A vs. C and A vs. D groups (Bonferroni $p < 0.0001$ for all three intergroup comparisons) Mean PPD values were similar between other groups (B vs. C: $p = 1.000$, B vs. D: $p = 1.000$, C vs. D: $p = 1.000$). CAL was similar among three groups of patients with diagnosed periodontitis (Table 1). Kappa scores were 0.5–0.6 for inter-rater agreement.

As expected, FPG level was statistically different between all groups, the highest being in the group of subjects with poorly regulated blood glucose level (Mann-Whitney U Test, $p < 0.05$ for all comparisons) (Table 1). HbA1C level was highest in group D, followed by group C. It differed between all groups except A vs. B (Mann-Whitney U Test, $p = 0.903$) (Table 1). Results of hematological measurements for Hgb, RBC, MCV and MCH were similar among groups (Kruskal-Wallis Test, $p > 0.05$ for all parameters). HCT was statistically different only between groups A and B (Mann-Whitney U Test, $p = 0.011$). MCHC values differed between groups B and C (Mann-Whitney U Test, $p = 0.011$) and groups C and D (Mann-Whitney U Test, $p = 0.016$).

Table 1. Demographic records, blood count and biochemical analysis data in groups.

	Groups				p value
	A (N = 36)	B (N = 42)	C (N = 28)	D (N = 40)	
Socio-demographic data					
Gender N(%) m / f	15(41.7) / 21(58.3)	17(40.5) / 25(59.5)	15(53.6) / 13(46.4)	26(65.0) / 14(35.0)	†.099
Age (x±SD)	43±4	48±11	48±7	47±7	‡.053
Smokers N (%)	8(22.2)	16(38.1)	9(32.1)	10(25.0)	†.408
Blood type sistem N (%)	O	5 (13.9)	8 (19.0)	6 (21.4)	‡.549
	A/B/AB	31 (86.1)	34 (81.0)	22 (78.6)	
Clinical periodontal parameters (presented as Mean±SD)					
Teeth number	27.58±1.422	20.36±5.28	16.89±4.646	14.85±3.899	§ < .0001*
PI	0.86±0.468	2.38±3.737	2.68±0.450	2.24±0.751	§ < .0001*
BOP (%)	39.58±19.099	61.24±24.390	62.58±25.370	64.40±28.888	‡ < .0001*
PPD (mm)	2.02±0.513	2.93±0.871	2.87±0.903	2.71±0.757	‡ < .0001*
CAL (mm)	NA	3.37±1.964	3.87±1.929	4.13±2.103	‡.233
Additional (biochemical and hematological) measurements (presented as Mean±SD)					
FPG (mmol/L)	4.64±0.534	4.97±0.578	7.59±1.824	11.24±4.061	§ < .0001*
HbA _{1c} (%)	4.80±0.607	4.82±0.561	7.10±0.566	10.81±1.357	§ < .0001*
Hgb (g/l)	137.08±10.308	114.81±52.717	139.75±11.670	137.19±14.731	§.355
RBC (10 ¹² /l)	4.61±0.474	4.31±1.197	4.62±0.375	4.69±0.601	§.458
MVC (fl)	89.15±9.299	91.82±9.857	89.15±3.620	88.78±9.686	§.567
HCT (l/l)	0.409±0.054	0.690±0.595	0.420±0.034	0.415±0.064	§.045*
MCH (pg)	29.91±2.565	28.82±2.747	30.27±1.489	29.43±2.511	‡.074
MCHC (g/l)	337.14±31.198	316.45±40.295	339.39±15.908	334.28±39.350	§.021*

X = mean; SD = standard deviation; m/f = males/females; PI = Silness Loe Plaque Index; BOP = bleeding on probing index; PPD = Probing Pocket Depth; CAL = Clinical Attachment Loss; FPG = Fasting plasma glucose level; HbA_{1c} = glycolysated hemoglobin; Hgb = Hemoglobin; RBC = Red Blood Count; MCV = Mean Corpuscular Volumen; HCT = Hematocrit; MCH = Mean Cell Hemoglobin; MCHC = mean corpuscular hemoglobin concentration

† Chi Square Test

‡ANOVA

§Kruskall-Wallis Test

*results with statistical significance

<https://doi.org/10.1371/journal.pone.0210527.t001>

Microbiological (primary) outcome variables

Candida spp. was detected on the tongue of 27.4% of subjects. It was more prevalent in diabetics with poor glycoregulation (47.5%) than in groups A (22.2%), B (16.7%) or C (21.5%) (Fig 2). Subjects with T2D (C+D = 25) had a significantly higher prevalence of *Candida* spp. detection (37.3%), than healthy subjects (A+B = 15 (19.5%)) (p = 0.013). The frequency of isolation of more than one *Candida* species (mixed isolation) per sample was higher in group D than in group A (p = 0.002) and group B (p = 0.018) (Table 2). The most frequently isolated species in this study was *C. albicans*—detected in all positive samples (27.4%), followed by *C. glabrata* (6.85%) and *C. tropicalis* (1.37%). In the samples of mixed infection of tongue, *Candida albicans* was the predominant species, presenting 75.83±19.287% (mean±SD) of isolates per sample. Even though there was no statistically significant difference in the amount of detected yeasts on tongue among groups (p = 0.331), it is interesting that group A presented only rare growth, while dense growth was detected only in group D.

In the samples obtained by sterile periodontal curette, 23.97% were positive (Fig 2). The frequency of *Candida* spp. isolation was higher in group D than in group B and group C. The

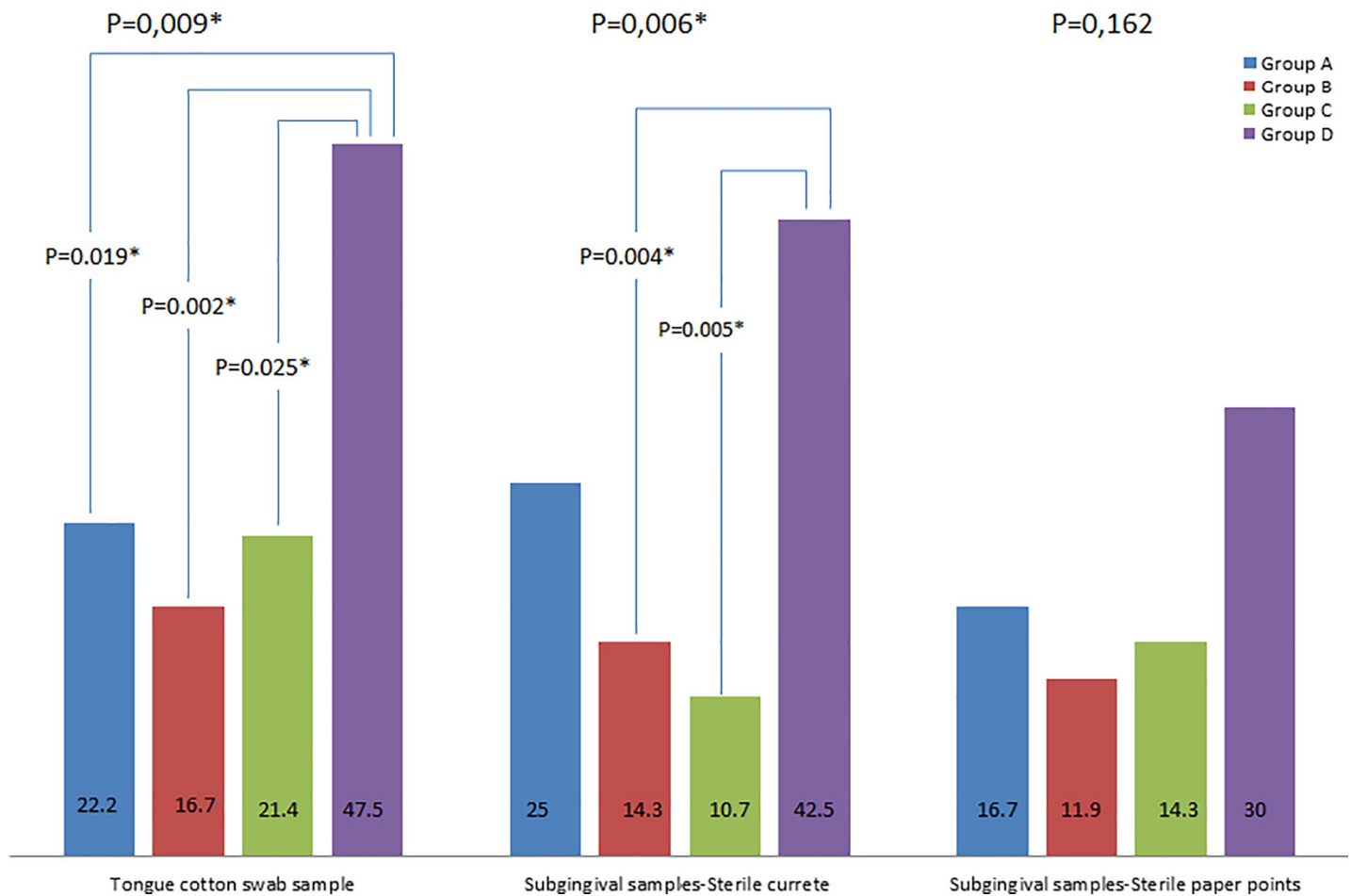


Fig 2. Positive findings of *Candida* spp. on tongue and in subgingival areas (presented as %). * presents statistical significant difference ($p < 0.05$).

<https://doi.org/10.1371/journal.pone.0210527.g002>

most frequently isolated species was *C. albicans* (23.97%), followed by *C. glabrata* (5.48%). The frequency of mixed isolation of *Candida* spp. was similar between the groups ($p = 0.152$) (Table 2). In the samples of mixed infection, *Candida albicans* was the predominant species, presenting $76.25 \pm 19.226\%$ (mean \pm SD) of isolates per sample.

Candida spp. was isolated in 18.49% of subgingival samples collected by sterile paper points (Fig 2). The most frequently isolated species was *C. albicans* (18.49%), followed by *C. glabrata* (4.79%) and *C. tropicalis* (0.80%). In the mixed infection of paper point's samples, *Candida albicans* was the predominant species, presenting $75.72 \pm 20.702\%$ (mean \pm SD) of isolates per sample. The frequency of mixed isolation of *Candida* spp. was similar between the groups ($p = 0.167$) (Table 2).

The presence of *C. albicans* in subgingival areas detected by either sampling method was 29.25%. Occurrence of *Candida* spp. in group D was significantly higher than in group A ($p = 0.018$), group B ($p = 0.023$) and group C ($p = 0.005$). Within each group, there was no difference in clinical periodontal parameters (PI, BOP, PPD, CAL) between patients with and without *Candida* spp. ($p > 0.05$ for all comparisons). Frequency of subgingival *Candida* detection differed according to sampling technique ($p < 0.0001$).

The number of subgingival CFU/ml did not differ between groups (Kruskal-Wallis, $p = 0.290$). The highest values for CFU/ml were detected in group B (1290.00 ± 1281.77), followed by group C (875.48 ± 485.63), group D (772.98 ± 659.43) and group A (478.57 ± 654.38) (expressed as mean \pm SD).

Table 2. *Candida* spp. finding on tongue and in subgingival areas sampled by sterile periodontal curette and sterile paper point.

Tongue samples (sterile cotton swab)					
	Group A (N = 36)	Group B (N = 42)	Group C (N = 28)	Group D (N = 40)	P value
Negative yeast finding N (%)	28 (77.8%)	35 (83.3%)	22 (78.5)	21 (52.5)	†0.009*
Positive yeast finding N (%)	8 (22.2%)	7 (16.7%)	6 (21.5)	19 (47.5)	
Non albicans finding: N (%)	0 (0)	1 (2.4%)	2 (7.1)	9(22.5)	
Sterile currette sample					
	Group A	Group B	Group C	Group D	P value
Negative yeast finding N (%)	27 (75%)	36 (87%)	25 (89%)	23 (57%)	†0.007*
Positive yeast finding N (%)	9 (25%)	6 (13%)	3 (11%)	17 (43)	
Non albicans finding: N (%)	1 (2.8)	1 (2.4)	1 (3.6)	5(12.5)	
Sterile paper points samples					
	Group A	Group B	Group C	Group D	P value
Negative yeast finding N (%)	30 (83%)	37 (88%)	24 (85%)	28 (70%)	†0.162
Positive yeast finding N (%)	6 (17%)	5 (12)	12 (30%)	12 (30%)	
Non albicans finding: N (%)	0 (0)	1 (2.4)	2 (7.1)	4 (10.0)	

† Chi Square Test

*results with statistical significance

<https://doi.org/10.1371/journal.pone.0210527.t002>

There were 15.07% of cases where yeasts were not detected on the tongue, but were found in the subgingival area. The frequencies of such findings were similar among groups ($p = 0.846$): A = 4/36 (11.1%), B = 6/42 (14.3%), C = 5/28 (17.9%), D = 7/40 (17.5%). Only 2.1% of subjects had different species of *Candida* present on tongue and in subgingival area, distributed similarly in groups B, C and D. On the other hand, only in 13.7% of subjects, both samples were positive.

Logistic regression analysis of the relationship among *Candida* carriage risk factor and prevalence of *Candida* spp. on tongue and in subgingival sites

Using logistic regression model, age, gender, blood type, smoking habits, presence of PD, presence of T2D, mentioned hematological and biochemical parameters were analyzed as potential predictors of *Candida* presence. FPG (OR = 1.138, CI = 1.031–1.256, $p = 0.011$) and HbA1c (OR = 1.273, CI = 1.112–1.459, $p < 0.0001$) have been shown as significant confounders. Only HbA1c (OR = 1.242, CI = 1.011–1.525, $p = 0.039$) was an independent predictor of the presence of *Candida* spp. using the multivariate logistic regression model.

The univariate logistic regression model was also applied in order to identify parameters that could predict positive detection of *Candida* spp. in the subgingival areas. Age, gender, blood type, smoking habits, presence of periodontitis or T2D, measured hematological and biochemical parameters and periodontal clinical parameters (PI, PPD and CAL) were analyzed as potential predictors. Although FPG (OR = 1.103, CI = 1.002–1.215, $p = 0.045$), Hb1c (OR = 1.242, CI = 1.087–1.415, $p = 0.001$) and RDW (OR = 1.219, CI = 1.009–1.474, $p = 0.040$) were confounders in univariate logistic regression analysis, only HbA1c was the confounder in multivariate regression model (OR = 1.290, CI = 1.062–1.567, $p = 0.010$).

Discussion

Although isolation of *Candida* spp. from the oral cavity does not mean disease per se, opinions about the role of yeasts isolated from periodontal pockets vary [10, 11]. Diabetic patients are

more susceptible to fungal carriage and infections [1, 18], as well as periodontitis commonly treated by antibiotics as adjuvant periodontal therapy. Therefore, the presence and role of yeasts in pathogenesis of periodontitis in immunocompromised subjects should be clarified.

The prevalence of oral yeasts in this study was determined using sterile cotton swabs of the middle and posterior thirds of the tongue midline, as it has been proven that *Candida* spp. most commonly inhabits these regions in the oral cavity [6]. This prevalence in our study was the highest in diabetics with poor glycoregulation, followed by diabetics with satisfactory glycoregulation, which is in concurrence with previously conducted studies [3, 19]. It is interesting that we found that the prevalence of *Candida* spp. on the tongue of diabetics with satisfactory glycoregulation was similar to the prevalence in both groups of healthy subjects. Bader et al. similarly concluded that good control of blood glucose level is the best protection against *Candida* infection [19]. Additionally, among a number of examined factors in this study, only HbA1c- parameter of glycoregulation has been proven to be an independent risk factor for positive yeast detection on the tongue. The prevalence of *Candida* spp. on the tongue in healthy patients is similar to another study exploring the frequency of this yeast on tongue of healthy subjects in Serbia [20]. Similar to the findings of Zomorodian et al. [3], *C. albicans* was the most frequent found species, followed by *C. glabrata*, and *C. tropicalis*- the last one isolated only at diabetics. Recovery of non- albicans species is important because of their high level of resistance to antifungal drugs [21].

Overall prevalence of subgingival *Candida* spp. was 29.25%. There is no standardized methodology for sampling the subgingival plaque, so in this study collection of subgingival biofilm was done by two commonly used techniques: paper points followed by sterile periodontal curette. Using both methodologies, we demonstrated that there is difference in efficiency of sampling methods. These results should encourage researchers to elucidate optimal methodology for sampling subgingival plaque as our research group started [22]. It is important to emphasize that our previously conducted study has been done at another study population. Results of this study showed higher prevalence of *Candida* spp. in subgingival areas of poorly regulated diabetic subjects than in groups A, B and C. Additionally, our results showed that HbA1c is the only independent predictor for detection of subgingival *Candida* spp. These results are in agreement with already mentioned data about prevention of *Candida* infection by good glycoregulation [13, 19]. Other studies showed occurrence of subgingival *Candida* spp. [11] in 10% of subjects with periodontal health, 30–44% in periodontitis patients [11, 14], and 35.2% to 52% in diabetics [13, 14]. In this study, subjects with periodontal health showed higher prevalence of subgingival yeasts (22.2%), but on the other hand periodontitis patients showed lower prevalence than other studies (14.6%). These surprisingly uncommon results may be explained by proofs about *in vitro* inhibitory effects of cariogenic (e.g. *Streptococcus mutans*) and periopathogenic (eg. *Prevotella intermedia*) bacteria on the growth of *Candida* spp. within biofilms [23], and oral microbiome interactions between *Candida* and other residents of oral mucosa [5]. Although we did not determine the presence of other microorganisms in subgingival areas, it can be assumed that there are more periopathogenic microorganisms in periodontal pockets than in the gingival sulci. The majority of studies did not divide patients according to glycoregulation when examining the prevalence of yeasts. The group with satisfactory glycoregulation showed similar prevalence to patients with periodontitis only, while subjects with poorly regulated blood glucose level showed higher prevalence. Although there are few of studies analyzing the presence of yeasts in subgingival areas, to the best of our knowledge, our study is of unique design. We considered clinical periodontal status in subjects without metabolic disorders and on the other hand, in subjects with T2D and CP, we considered glycoregulation. Even though we planned a group of patients with T2D and

clinically healthy periodontium, we could not find any subjects who fitted these criteria, so we omitted this group.

Comparable to other studies, *C. albicans* was the most frequently isolated species [11, 13], but unlike these studies we did not find *C. dubliniensis* in subgingival samples. The frequency of *C. glabrata* was similar to mentioned studies. *C. tropicalis* was isolated only in T2D patients, which is in agreement with the fact that this species is commonly isolated from immunocompromised patients [24] and not from healthy ones [3]. Some researches consider *C. tropicalis* as the third most commonly isolated non-*albicans* species, which is associated with higher mortality than other non-*albicans* species [24], and has ability to develop rapid resistance to fluconazole [25], so recognition of potential reservoirs of this species on clinically healthy tongue or in periodontal pockets could be useful. Similarly, *C. glabrata* is also considered as second or third most common isolate of *Candida* spp. [26], and is increasingly connected with mucosal and general infections, especially in diabetics [27]. In our study, it has also been found in healthy subjects with periodontitis, but even in one sample from clinically healthy periodontium (isolated only by sterile curette). Since periodontitis itself has been defined as a state of disturbed cellular and humoral immune local response [28], it can be expected for *C. glabrata* to be found in periodontal pockets. It should be stressed that colonization does not necessarily indicate candidiasis, so *C. glabrata* in gingival sulcus can be observed as transient member or possibly a reservoir of yeasts. Also, these two species have been already isolated from oral cavities of healthy persons, but without taking into consideration the periodontal status [29].

The subgingival area as a reservoir of yeasts should be taken into consideration seriously, since in our study 15.7% of subjects exhibited *Candida* spp. only in subgingival biofilm, similar to the findings of Hammad et al [30]. Patients included in this study were without symptoms and medical history of candidiasis treatment. Also, in clinical praxis when candidiasis is suspected to be present, oral swabs are taken from the oral mucosa, but not from the subgingival area. Additionally, *Candida* spp. in subgingival areas is more resistant because it is always present within biofilms, and additionally these sites are inaccessible to antifungal drugs [31]. The presence of *Candida* spp. in subgingival area is important because biofilms increase resistance to conventional antifungal agents [32, 33]. In the case of inflamed periodontal tissue which means ulcerated epithelium of periodontal pocket, the entrance of microorganisms into the bloodstream is enabled. In cases of immunocompromised patients, this could potentially be life-threatening [34]. Since oral *Candida* spp. carriage may be influenced by many factors non-related to diabetes (gender, smoking, medications, saliva, oral site, blood type (O vs. A, AB and B blood type) [35]), we analyzed various possible factors which can be potentially confounders for *Candida* spp. carriage. To the best of our knowledge, this is the first study that took into consideration almost all factors associated with yeasts. As it is mentioned earlier, only HbA1c was predictor for the presence of *Candida*, but with a low risk ratio. Subgingival *Candida* carriage has already been connected with glycoregulation [13, 30], but results of the connection between HbA1c level and yeast detection on tongue varies [3, 19, 30]. Evidence about the role of smoking and gender on *Candida* spp. colonization and infections differ between studies [12, [20, 36, 37]. In our study, these parameters, similarly distributed between the groups, did not influence the *Candida* presence.

We did not find a relationship between clinical periodontal parameters and the presence of subgingival yeasts. Although it has been proven that *Candida* spp. colonization is affected by hygiene habits [38], some studies also did not find a correlation between amount of dental biofilm or gingival status with the presence of subgingival yeasts [30, 39].

Even though there is a lot of evidence about the presence of yeasts in periodontal pockets, their role in pathogenesis of periodontitis is still to be elucidated. Detection of *Candida* spp. in

subgingival biofilm is only the first step for connecting yeast and periodontitis. As mentioned earlier, this presence could be transient [10], so longitudinal rather than cross sectional studies are needed for making accurate conclusions. Cross sectional design of the study, along with only one site (the deepest periodontal pocket) of subgingival biofilm collection are the main limitations of this study. Likewise, immunological response around different species and morphological forms of *Candida* spp. should be examined, but also taking into consideration all factors that could potentially affect the presence and quantity of yeasts. Variation of *Candida* carriage depending on the geographical region and/or patient group [40] are additional problems. In some countries, even non-albicans forms could be more frequently isolated than *C. albicans* [40]. In Serbia, there are not a sufficient number of studies examining the presence of different *Candida* species. A study examining prevalence of *C. albicans* and non-albicans species at oral lesions [41] and examining their extra oral prevalence [42] showed also highest prevalence of *C. albicans* [43].

As mentioned, besides the potential role in periodontitis, subgingival areas could be a potential reservoir and focus of yeasts, so the new approaches to the therapy of oral candidiasis, including obligatory scaling and root planing should be considered.

Conclusions

- Presence of *Candida* spp. on the tongue is higher in diabetic with poor glycoregulation and is only influenced by HbA1c serum level (glycoregulation).
- Presence of subgingival *Candida* spp. has not been influenced by parameters of oral hygiene or clinical periodontal parameters, but was influenced by HbA1c serum level.
- The presence of *Candida* spp. in subgingival areas (even when they are not found on the tongue) indicate that subgingival biofilms could be a potential reservoir of these microorganisms.

Supporting information

S1 File. Questionnaire used in the study (English).
(PDF)

S2 File. Questionnaire used in the study (Serbian).
(PDF)

S3 File. All collected data.
(SAV)

Author Contributions

Conceptualization: Sanja Matic Petrovic, Milena Radunovic, Valentina Arsic Arsenijevic, Ana Pucar.

Formal analysis: Jovana Kuzmanovic Pfcicer.

Funding acquisition: Ana Pucar.

Investigation: Sanja Matic Petrovic, Milena Radunovic, Milena Barac.

Methodology: Sanja Matic Petrovic, Milena Radunovic, Ana Pucar.

Project administration: Dusan Pavlica, Valentina Arsic Arsenijevic, Ana Pucar.

Resources: Sanja Matic Petrovic, Milena Radunovic.

Supervision: Ana Pucar.

Validation: Milena Barac.

Visualization: Sanja Matic Petrovic, Milena Radunovic.

Writing – original draft: Sanja Matic Petrovic, Milena Radunovic.

Writing – review & editing: Jovana Kuzmanovic Pficer, Dusan Pavlica.

References

1. Guggenheimer J, Moore PA, Rossie K, Myers D, Mongelluzzo MB, Block HM, et al. Insulin-dependent diabetes mellitus and oral soft tissue pathologies. II. Prevalence and characteristics of *Candida* and candidal lesions. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*. 2000; 89(5):570–6. PMID: [10807713](#)
2. Løe H. Periodontal Disease: The sixth complication of diabetes mellitus. *Diabetes Care*. 1993; 16(1):329–34. PMID: [8422804](#)
3. Zomorodian K, Kavooosi F, Pishdad GR, Mehriar P, Ebrahimi H, Bandegani A, et al. Prevalence of oral *Candida* colonization in patients with diabetes mellitus. *Journal de Mycologie Médicale / Journal of Medical Mycology*. 2016; 26(2):103–10. <https://doi.org/10.1016/j.mycmed.2015.12.008> PMID: [26879707](#)
4. Papon N, Courdavault V, Clastre M, Bennett RJ. Emerging and Emerged Pathogenic *Candida* Species: Beyond the *Candida albicans* Paradigm. *PLOS Pathogens*. 2013; 9(9):e1003550. <https://doi.org/10.1371/journal.ppat.1003550> PMID: [24086128](#)
5. Mukherjee PK, Chandra J, Retuerto M, Sikaroodi M, Brown RE, Jurevic R, et al. Oral Mycobiome Analysis of HIV-Infected Patients: Identification of *Pichia* as an Antagonist of Opportunistic Fungi. *PLOS Pathogens*. 2014; 10(3):e1003996. <https://doi.org/10.1371/journal.ppat.1003996> PMID: [24626467](#)
6. Arendorf TM, Walker DM. The prevalence and intra-oral distribution of *Candida albicans* in man. *Archives of Oral Biology*. 1980; 25(1):1–10. PMID: [6996654](#)
7. Colombo APV, Magalhães CB, Hartenbach FARR, do Souto RM, da Silva-Boghossian CM. Periodontal-disease-associated biofilm: A reservoir for pathogens of medical importance. *Microbial pathogenesis*. 2016; 94:27–34. <https://doi.org/10.1016/j.micpath.2015.09.009>
8. Grenier G, Gagnon G, Grenier D. Detection of herpetic viruses in gingival crevicular fluid of patients suffering from periodontal diseases: prevalence and effect of treatment. *Oral Microbiology and Immunology*. 2009; 24(6):506–9. <https://doi.org/10.1111/j.1399-302X.2009.00542.x> PMID: [19832804](#)
9. Järvensivu A, Hietanen J, Rautemaa R, Sorsa T, Richardson M. *Candida* yeasts in chronic periodontitis tissues and subgingival microbial biofilms in vivo. *Oral Diseases*. 2004; 10(2):106–12. PMID: [14996281](#)
10. Hintao J, Teanpaisan R, Chongsuvivatwong V, Ratarasan C, Dahlén G. The microbiological profiles of saliva, supragingival and subgingival plaque and dental caries in adults with and without type 2 diabetes mellitus. *Oral Microbiology and Immunology*. 2007; 22(3):175–81. <https://doi.org/10.1111/j.1399-302X.2007.00341.x> PMID: [17488443](#)
11. Urzúa B, Hermosilla G, Gamonal J, Morales-Bozo I, Canals M, Barahona S, et al. Yeast diversity in the oral microbiota of subjects with periodontitis: *Candida albicans* and *Candida dubliniensis* colonize the periodontal pockets. *Medical mycology*. 2008; 46(8):783–93. <https://doi.org/10.1080/13693780802060899> PMID: [18608938](#)
12. Yuan K, Chang C-j, Hsu P-c, Sun HS, Tseng C-c, Wang J-r. Detection of putative periodontal pathogens in non-insulin-dependent diabetes mellitus and non-diabetes mellitus by polymerase chain reaction. *Journal of periodontal research*. 2001; 36(1):18–24. PMID: [11246700](#)
13. Al Mubarak S, Robert AA, Baskaradoss JK, Al-Zoman K, Al Sohail A, Alsuwyyed A, et al. The prevalence of oral *Candida* infections in periodontitis patients with type 2 diabetes mellitus. *Journal of Infection and Public Health*. 2013; 6(4):296–301. <https://doi.org/10.1016/j.jiph.2012.12.007> PMID: [23806705](#)
14. Melton JJ, Redding SW, Kirkpatrick WR, Reasner CA, Ocampo GL, Venkatesh A, et al. Recovery of *Candida dubliniensis* and other *Candida* species from the oral cavity of subjects with periodontitis who had well-controlled and poorly controlled type 2 diabetes: a pilot study. *Special Care in Dentistry*. 2010; 30(6):230–4. <https://doi.org/10.1111/j.1754-4505.2010.00159.x> PMID: [21044102](#)
15. 1999 International Workshop for a Classification of Periodontal Diseases and Conditions. Papers. Oak Brook, Illinois, October 30- November 2, 1999. *Ann Periodontol* 1999; 4:i, 1–112
16. Association AD. Standards of Medical Care in Diabetes—2013. *Diabetes Care*. 2013; 36(Supplement 1):S11–S66.

17. Marinho SA, Teixeira AB, Santos OS, Cazanova RF, Ferreira CAS, Cherubini K, et al. Identification of *Candida* spp. by phenotypic tests and PCR. *Brazilian Journal of Microbiology*. 2010; 41:286–94. <https://doi.org/10.1590/S1517-83822010000200004> PMID: 24031493
18. Cullinan M, Ford P, Seymour G. Periodontal disease and systemic health: current status. *Australian Dental Journal*. 2009; 54(s1):S62–S9.
19. Bader M, Hinthorn D, Lai S, Ellerbeck E. Hyperglycaemia and mortality of diabetic patients with candidaemia. *Diabetic medicine*. 2005; 22(9):1252–7. <https://doi.org/10.1111/j.1464-5491.2005.01626.x> PMID: 16108857
20. Cankovic M, Bokor-Bratic M, Cankovic D. Oral fungal and bacterial infection in smokers. *HealthMed*. 2011; 5(6):1695–700.
21. González GM, Elizondo M, Ayala J. Trends in species distribution and susceptibility of bloodstream isolates of *Candida* collected in Monterrey, Mexico, to seven antifungal agents: results of a 3-year (2004 to 2007) surveillance study. *Journal of clinical microbiology*. 2008; 46(9):2902–5. <https://doi.org/10.1128/JCM.00937-08> PMID: 18632907
22. Matic Petrovic S, Cimbaljevic M, Radunovic M, Kuzmaanovic Pficer J, Jotic A, Pucar A. Detection and sampling methods for isolation of *Candida* spp. from oral cavities in diabetics and non-diabetics. *Brazilian Oral Research*. 2015; 29:1–7.
23. Thein ZM, Samaranyake YH, Samaranyake LP. Effect of oral bacteria on growth and survival of *Candida albicans* biofilms. *Archives of Oral Biology*. 2006; 51(8):672–80. <https://doi.org/10.1016/j.archoralbio.2006.02.005> PMID: 16620775
24. Colombo AL, Guimarães T, Silva LR, de Almeida Monfardini LP, Cunha AKB, Rady P, et al. Prospective observational study of candidemia in Sao Paulo, Brazil: incidence rate, epidemiology, and predictors of mortality. *Infection Control & Hospital Epidemiology*. 2007; 28(05):570–6.
25. Yang Y-L, Ho Y-A, Cheng H-H, Ho M, Lo H-J. Susceptibilities of *Candida* species to amphotericin B and fluconazole: the emergence of fluconazole resistance in *Candida tropicalis*. *Infection Control & Hospital Epidemiology*. 2004; 25(01):60–4.
26. Sinnott JT, Cullison JP, Sweeney MP. *Candida* (*Torulopsis*) *glabrata*. *Infection Control*. 1987; 8(08):334–6.
27. Hitchcock C, Pye G, Troke P, Johnson E, Warnock D. Fluconazole resistance in *Candida glabrata*. *Antimicrobial agents and chemotherapy*. 1993; 37(9):1962–5. PMID: 8239613
28. Murray PA. Periodontal diseases in patients infected by human immunodeficiency virus. *Periodontology* 2000. 1994; 6(1):50–67.
29. Sánchez-Vargas LO, Ortiz-López NG, Villar M, Moragues MD, Aguirre JM, Cashat-Cruz M, et al. Point prevalence, microbiology and antifungal susceptibility patterns of oral *Candida* isolates colonizing or infecting Mexican HIV/AIDS patients and healthy persons. *Revista iberoamericana de micología*. 2005; 22(2):83. PMID: 16107165
30. Hammad MM, Darwazeh AMG, Idrees MM. The effect of glycemic control on *Candida* colonization of the tongue and the subgingival plaque in patients with type II diabetes and periodontitis. *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*. 2013; 116(3):321–6. <https://doi.org/10.1016/j.oooo.2013.05.013> PMID: 23953417
31. Oliveira LFd, Jorge AOC, Santos SSF. In vitro minocycline activity on superinfecting microorganisms isolated from chronic periodontitis patients. *Brazilian oral research*. 2006; 20(3):202–6. PMID: 17119701
32. Al-Attas S, Amro S. Candidal colonization, strain diversity, and antifungal susceptibility among adult diabetic patients. *Annals of Saudi medicine*. 2010; 30(2):101. <https://doi.org/10.4103/0256-4947.60514> PMID: 20220258
33. Mallick EM, Bennett RJ. Sensing of the Microbial Neighborhood by *Candida albicans*. *PLOS Pathogens*. 2013; 9(10):e1003661. <https://doi.org/10.1371/journal.ppat.1003661> PMID: 24204254
34. Sardi JCO, Almeida AMF, Mendes Giannini MJS. New antimicrobial therapies used against fungi present in subgingival sites—a brief review. *Archives of oral biology*. 2011; 56(10):951–9. <https://doi.org/10.1016/j.archoralbio.2011.03.007> PMID: 21676377
35. Burford-Mason A, Weber J, Willoughby J. Oral carriage of *Candida albicans*, ABO blood group and secretor status in healthy subjects. *Journal of medical and veterinary mycology*. 1988; 26(1):49–56. PMID: 3288742
36. Darwazeh A, Al-Dwairi Z, Al-Zwairi A. The relationship between tobacco smoking and oral colonization with *Candida* species. *J Contemp Dent Pract*. 2010; 11(3):017–24. PMID: 20461320
37. Rasool S, Siar C, Ng K. Oral candidal species among smokers and non-smokers. *Journal of the College of Physicians and Surgeons—Pakistan: JCPSP*. 2005; 15(11):679–82. PMID: 16300700

38. Budtz-Jørgensen E, Mojon P, Rentsch A, Deslauriers N. Effects of an oral health program on the occurrence of oral candidosis in a long-term care facility. *Community Dentistry and Oral Epidemiology*. 2000; 28(2):141–9. PMID: [10730723](https://pubmed.ncbi.nlm.nih.gov/10730723/)
39. Darwazeh A-G, Hammad MM, Al-Jamaei AA. The relationship between oral hygiene and oral colonization with *Candida* species in healthy adult subjects*. *International Journal of Dental Hygiene*. 2010; 8(2):128–33. <https://doi.org/10.1111/j.1601-5037.2009.00407.x> PMID: [20522136](https://pubmed.ncbi.nlm.nih.gov/20522136/)
40. Colombo A, Perfect J, DiNubile M, Bartizal K, Motyl M, Hicks P, et al. Global distribution and outcomes for *Candida* species causing invasive candidiasis: results from an international randomized double-blind study of caspofungin versus amphotericin B for the treatment of invasive candidiasis. *European Journal of Clinical Microbiology and Infectious Diseases*. 2003; 22(8):470–4. <https://doi.org/10.1007/s10096-003-0973-8> PMID: [12884068](https://pubmed.ncbi.nlm.nih.gov/12884068/)
41. Bokor-Bratic M, Cankovic M, Dragic N. Unstimulated whole salivary flow rate and anxiolytics intake are independently associated with oral *Candida* infection in patients with oral lichen planus. *European Journal of Oral Sciences*. 2013; 121(5):427–33. <https://doi.org/10.1111/eos.12073> PMID: [24028590](https://pubmed.ncbi.nlm.nih.gov/24028590/)
42. Otašević S, Barac A, Pekmezovic M, Tasic S, Ignjatović A, Momčilović S, et al. The prevalence of *Candida* onychomycosis in Southeastern Serbia from 2011 to 2015. *Mycoses*. 2016; 59(3):167–72. <https://doi.org/10.1111/myc.12448> PMID: [26710983](https://pubmed.ncbi.nlm.nih.gov/26710983/)
43. Perić M, Živković R, Lemić AM, Radunović M, Miličić B, Arsenijević VA. The severity of denture stomatitis as related to risk factors and different *Candida* spp. *Oral surgery, oral medicine, oral pathology and oral radiology*. 2018; 126(1):41–47.